

## Troubleshooting With You



### Quantitative Real-time Polymerase Chain Reaction (qRT-PCR)

**Fatemeh Mehrpooyan\***

MSc., Genetic and Molecular Biology, Faculty of Science, University of Malaya, Kuala Lumpur, Malaysia.

\*sahar\_mehr8261@yahoo.com

- **What is Real Time PCR?**

In molecular biology, real-time polymerase chain reaction, also called quantitative real time polymerase chain reaction (qPCR) is a laboratory technique based on the PCR, which is used to amplify and quantify a targeted DNA molecule at the same time. For one or more specific sequences in a DNA sample, real-time PCR enables both detection and quantification. The quantity can be either an absolute number of copies or a relative amount when normalized to additional normalizing endogenous controls. Also, real-time PCR is combined with reverse transcription to quantify mRNA and non-coding RNA in cells or tissues.

- **How does it work?**

Reverse transcription is done using the High Capacity cDNA Reverse transcription kit (Applied Biosystems, USA). Each 20  $\mu$ l reaction mixture contains 2  $\mu$ g of total RNA, 4 mM of 25X dNTP mix (100 mM), 2  $\mu$ l (1X) of 10X RT Random primers, 2  $\mu$ l (1X) of 10X RT buffer, 1U/ $\mu$ l of 40 U/ml Ambion RNase Inhibitor, 1  $\mu$ l (50 U) of 50 U/ml MultiScribe™ Reverse transcriptase topped up with DEPC treated water. The thermal cycling profile is as follows: 25 °C 10 min 37 °C 120 min 85 °C 5 seconds and hold 4 °C.

Quantitative PCR amplification will be carried out using Power SYBR- Green PCR Master Mix kit (Applied Biosystems, USA). For each PCR, 12.5  $\mu$ l (1X) 2XSYBR Green Power Master Mix will be mixed with 500 nM of each forward and reverse primer solutions (10  $\mu$ M), 100 ng cDNA template topped up with autoclaved dH<sub>2</sub>O to a final volume of 25  $\mu$ l. The PCR Master mixes will be prepared using both  $\beta$ -actin endogenous control strips (the most stable control gene) as well as desired target RNA primers for the sample strips. The master mix containing endogenous primers will be added into the endogenous control strips while the master mix containing the main sample primers will be added into the sample strips. cDNA and nuclease free water will then be added to the specific strips. For each sample at least three replicates should be performed. No Template Control (NTC) tubes also will be prepared using both endogenous and sample primers to ensure that there will not be any master mix or/and primer contamination. Then the optical 8 tubes strips will be loaded into the Applied Biosystem 7500 Real-Time PCR system and the SDS 1.3.1 (Sequence Detection Software) is used to create a relative quantification (ddCt) plate. The run then will be started. The amplification program is: 95 °C for 10 minutes, 40 cycles of 95 °C for 15 seconds and finally 60 °C for 1 minute.

After the running of the program, the SDS software will be again used to generate a dissociation curve and later a relative quantification (ddCt) study will be created to analyze the results. The SDS software

on the Applied Biosystem 7500 Real-Time PCR system automatically uses the equation  $2^{-UUCT}$  to calculate the expression level of the pre-miRNA relative to the endogenous control.

- ***What is it used for?***

There are several applications for real-time PCR in laboratories. It is usually used for both diagnostic and basic research. Real-time PCR is mostly used to provide quantitative measurements of gene transcription. It may be used in recognizing how the expression of a particular gene changes over the time, e.g., in the response of tissue and cell cultures to an administration of a pharmacological agent, progression of cell differentiation, or in response to alterations in environmental conditions.

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[farzamisepehr@iau-saveh.ac.ir](mailto:farzamisepehr@iau-saveh.ac.ir)

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Department of Biology  
Faculty of Science  
Islamic Azad University, Gorgan Branch  
Gorgan, Iran

[ghorbanli@yahoo.com](mailto:ghorbanli@yahoo.com)

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[Eisvand.hr@iu.ac.ir](mailto:Eisvand.hr@iu.ac.ir)

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Assistant Professor  
Department of Biology, Faculty of Agriculture  
Islamic Azad University, Saveh Branch, Saveh, Iran

[farzamisepehr@iau-saveh.ac.ir](mailto:farzamisepehr@iau-saveh.ac.ir)

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Associate Professor  
Department of Horticultural science  
Islamic Azad University, Saveh Branch, Saveh, Iran

[pjmoradi@iau-saveh.ac.ir](mailto:pjmoradi@iau-saveh.ac.ir)



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## *Iranian Journal of Plant Physiology*

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**Ouyang, D., J. Bartholic and J. Selegean**, 2005. 'Assessing sediment loading from agricultural croplands in the great lakes basin'. *Journal of American Science*, 1 (2): 14-21.

### Books:

**Durbin, R., S. R. Eddy, A. Krogh and G. Mitchison**. 1999. *Biological Sequence Analysis: Probabilistic Models of Proteins and Nucleic Acids*. Cambridge: University Press.

### A chapter in a book:

**Leach, J.** 1993. 'Impacts of the zebra mussel (*Dreissena polymorpha*) on water quality and fish spawning reefs of Western Lake Erie'. In *Zebra Mussels: biology, impacts and control*. Nalepa, T. and D. Schloesser (Eds.). Ann Arbor, MI: Lewis Publishers, pp: 381-397.

### A Report:

**Makarewicz, J. C., T. Lewis and P. Bertram**. 1995. *Epilimnetic phytoplankton and zooplankton biomass and species composition in Lake Michigan 1983-1992*. U.S. EPA Great Lakes National Program, Chicago, IL. EPA 905-R-95-009.

### Conference proceedings:

**Stock, A.** 2004. 'Signal transduction in bacteria'. Proceedings of the 2004 Markey Scholars Conference, pp: 80-89.

### A thesis:

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