



Fractional purification and kinetic parameters (K_m and V_{max}) of polyphenol oxidase extracted from three segments of *Solanum melongenas* and *Musa sapientum* fruits

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Abstract

Polyphenol oxidase (PPO) was extracted from three segments of *Solanum melongenas* and *Musa sapientum* fruits and partially purified. The specific activity of PPO was measured at each purification step to ascertain level of enzyme purity. In all cases, PPO conformed to Michaelis-Menten kinetics, showing different values of kinetics parameters. Michaelis-Menten constant (PPO_{K_m}) of *S. melongenas* mid-section and anterior segments showed no significant difference ($p < 0.05$), whereas the posterior gave $PPO_{K_m} = 4.6 \pm 0.49$ mM ($p > 0.05$). Maximum PPO activity ($PPO_{V_{max}}$) was highest in the posterior segment: $PPO_{V_{max}} = 0.602 \pm 0.09$ U. Mid-section of *M. sapientum* exhibited the highest K_m value ($PPO_{K_m} = 5.8 \pm 0.69$ mM) compared with the anterior ($PPO_{K_m} = 3.9 \pm 0.69$ mM) ($p > 0.05$) and posterior $PPO_{K_m} = 4.9 \pm 0.11$ mM segments ($p < 0.05$). Overall, *M. sapientum* PPO_{K_m} values were relatively higher than those of *S. melongenas*. Posterior *S. melongenas* exhibited the highest $PPO_{V_{max}} = 0.602 \pm 0.09$ U, whereas the lowest value was registered in the anterior segment of *M. sapientum* $PPO_{V_{max}} = 0.234 \pm 0.09$ U. Substrate specificity for PPO (PPO_{V_{max}/K_m}) extracted from various segments of *S. melongenas* was in the increasing order of Mid-section > Posterior > Anterior, whereas that of *M. sapientum* was Mid-section > Anterior > Posterior. PPO_{V_{max}/K_m} between the two fruits showed strong positive correlation ($r = 0.862339$). Catechol was a better substrate for $PPO_{S. melongenas}$ than $PPO_{M. sapientum}$. The experimentally observed kinetic parameters of *S. melongenas* and *M. sapientum* signified the presence of PPO isoenzymes and non-uniform distribution of PPO in the two fruits.

Keywords: polyphenol oxidase, *Solanum melongenas*, *Musa sapientum*, kinetics parameters

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Introduction

Enzymatic browning is the discoloration that results from the action of a group of

enzymes collectively called polyphenol oxidases (PPO) (Broothaerts et al., 2000; Gouzi et al., 2010). The enzyme action is initiated by disruption of cell integrity and when the content of plastid and vacuole are mixed caused by senescence, wounding, or other tissue damage (Casado-Vela et al., 2005; Escobar et al., 2008). In

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addition, Thipyapong et al. (2004) suggested that PPO might have a role in the development of plant water stress and potential for photo-inhibition and photo-oxidative damage. Enzymatic browning reaction involves interaction of phenolic compounds with PPO in the presence of molecular oxygen (Kavrayan and Aydemir, 2001). PPO catalyzes two reactions including hydroxylation of monophenols to give *o*-diphenol (monophenol oxidase, cresolase tyrosinase activity EC. 1.14.18.1) (Klabunde et al., 1998; Fawzy, 2005) and oxidation of *o*-diphenol to *o*-quinones (diphenol oxidase, catecholase activity EC.1.10.3.1) (Mayer, 2006; Madani et al., 2011). The *o*-quinones readily polymerize and/or react with endogenous amino acids and proteins to form complex brown pigments (Klabunde et al., 1998; Casado-Vela et al., 2005; Prohp et al., 2009). Some of PPO substrates that occur naturally in fruits and vegetables, and are very suitable to enzymatic browning, include chlorogenic acid, catechin and epicatechin (Queiroz et al., 2008).

PPO is a copper (Cu^{2+}) containing metalloenzyme predominantly located in the chloroplast thylakoid membrane (Anthon and Barrett, 2002). The two atoms of Cu^{2+} are tightly ligated to three histidine residues of a polypeptide chain (Klabunde et al., 1998). The enzyme exists in isoforms (Barthet, 1997; Casado-Vela et al., 2005; Chikezie, 2006; Anderson et al., 2006; Escobar et al., 2008) and as zymogen (Gandia-Herrero et al., 2004; Sellés-Marchart et al., 2006). PPO activation can be achieved by variety of treatments such as urea (Okot-Kotber, 2002), et al., polyamines (Jimenez-Atienzar 1991), anionic detergents such as sodium dodecyl sulphate (SDS) (Santosh et al., 2006) and trypsin or proteinase K (Marques et al. 1994; Laveda et al., 2001). The state of PPO in plant tissues is approximately 85% met-PPO and 10-15% oxy-PPO forms and often isolated in the met-PPO form (Parkin, 2008).

The molecular weight of PPO extracted from different plant species have been reported by several authors. Probably due to partial proteolysis of the enzyme during its isolation, the molecular weight of plant PPO is very diverse and variable, e.g., *Eriobotrya japonica* Lindl: 59.2-61.2 kDa (Sellés-Marchart et al., 2006), *Brassica*

oleracea: 39 kDa (Fujita et al., 1995), *M. sapientum*: 62 kDa (Galeazzi et al., 1981), *Phaseolus vulgaris* L: 120 kDa (Beena and Gowda, 2000), *Malpighia glabra* L: 52 and 38 kDa (Kumar et al., 2008), *Manihot esculenta* C- isoenzymes PPO₁: 71.8 kDa and PPO₂: 69.6 kDa (Barthet, 1997) and *Brassica rapa*: 65 kDa (Nagai and Suzuki, 2001).

Solanum melongenas commonly referred to as garden egg in Nigeria and banana (*Musa sapientum*) are fruits widely grown as cash and food crops in the Tropics. Browning reaction of fresh-cut fruits and vegetables is a crucial and limiting factor determining the shelf life and acceptability of these products. Understanding the biochemical properties and kinetics of PPO is an imperative for applying control measures to mitigate this undesirable reaction.

Previous reports on the kinetic properties of PPO involved the study of enzyme extracts obtained from whole fruits and vegetables (Anthon and Barrett, 2002; Yagar and Sagiroglu, 2002; Sellés-Marchart et al., 2006; Chisari et al., 2007; Kumar et al., 2008; Queiroz et al., 2008; Prohp et al., 2009; Gouzi et al., 2010). Furthermore, there are reports on isoforms (Vámos-Vigyázó, 1981; Barthet, 1997; Casado-Vela et al., 2005; Chikezie, 2006; Altunkaya and Gökmen, 2011) and non-uniform distribution of PPO in plant systems (Qudsieh et al., 2002; Sirhindi, 2003). Therefore, these earlier reported kinetic properties of PPO extracted from whole fruits and vegetables probably did not represent the true kinetic features of the various PPOs in those plant specimen.

The present study seeks to measure two kinetic parameters, Michaelis Menten (K_m) and maximum velocity (V_{max}), of PPO extracted from the posterior, mid-section and anterior segments of *S. melongenas* and *M. sapientum* fruits. The study will give an insight into kinetic properties and, by extension, relative abundance / distribution of PPO in the three portions of the two fruits under investigation.

Materials and Methods

Collection and preparation of fruit samples

Fresh and disease free fruits of *S. melongenas* and *M. sapietum* were harvested from a private botanical garden in Umuoziri-Inyishi, Imo State, Nigeria between 17th -30th of July, 2012. The fruits were identified and authenticated by Dr. F. N. Mbagwu at the Herbarium in the Department of Plant Science and Biotechnology, Imo State University, Owerri, Nigeria. The two fruits were washed under continuous current of distilled water for 5 min and air dried at room temperature. The stalk (*S. melongenas*) and rind (*M. sapietum*) were removed manually. The samples were cut into three distinct segments: anterior, mid-section and posterior and stored at -4 °C until used for analyses.

Extraction and purification of PPO

Extraction and partial purification of PPO was according to the methods of Madani et al., (2011) with minor modifications. Ten grams (10 g) of the sample was homogenized in external ice bath, using Ultra-Turrax T25 (Janke and Kunkel, Staufen, Germany) homogenizer set in 80 mL of 0.1 M phosphate buffer (pH 6.8) containing 10 mM ascorbic acid for 180 sec at intervals of 60 sec. The homogenate was quickly squeezed through two layers of clean cheese cloth into a beaker kept in ice. The crude extract samples were centrifuged at 32000 *g* for 20 min at 4 °C. Solid ammonium sulphate (NH₄)₂SO₄ was added to the supernatant to obtain 80% (NH₄)₂SO₄ saturation and precipitated proteins were separated by centrifugation at 32000 *g* for 30 min at 4 °C. The precipitate was re-dissolved in 10 mL distilled water and ultra-filtered in a Millipore stirred cell with a 10-kDa membrane (Millipore 8050, Milan, Italy). The filtrate was dialyzed at 4 °C against distilled water for 24 h with 4 changes of the water during dialysis. The dialyzed sample constituted the partial purified PPO extract and was used as the enzyme source from the corresponding segments of the two fruits. Protein concentrations were determined by the method of Bradford, (1976) using bovine serum albumin as standard at $\lambda_{\max} = 595$ nm. One unit of PPO activity was defined as the amount of enzyme that causes an increase in absorbance of 0.001 mL⁻¹ min⁻¹ under the condition of the assay

(Oktay *et al.*, 1995). The procedure and measure of PPO purification is summarized in Table 1.

Determination of PPO activity

PPO activity was measured immediately after the extract was partially purified. Enzyme assay was according to the methods of Qudsieh et al. (2002) with minor modifications (Chikezie, 2006). Enzyme activity was determined by measuring the increase in absorbance at 540 nm using a spectrophotometer (U-2000 Hitachi, Japan) at 24 °C. The reaction mixture contained 3.5 mL of 0.20 M phosphate buffer (pH = 6.8), 1 mL of each serial dilutions of 12 - 0.75 mM catechol, and 0.5 mL of enzyme solution in a final volume of 5 mL. The mixture was quickly transferred into a cuvette and the change in absorbance was monitored at $\lambda_{\max} = 540$ nm at a regular interval of 30 sec. The rate of the reaction was calculated from the initial linear slope of activity curves.

Evaluation of kinetic constants

The K_m and V_{max} values of PPO were measured with the use of the Lineweaver– Burk (1/ V_o versus 1/[S] values) graphs (Lineweaver and Burk, 1934).

Statistical analysis

The experiments were designed in a completely randomized method and the collected data were analyzed by the analysis of variance procedure while treatment means were separated by the least significance difference (LSD) incorporated in the statistical analysis system (SAS) package of 9.1 version, (2006). The correlation coefficients between the results were determined with Microsoft Office Excel, 2010 version.

Results

The specific activity of PPO extracts, which was a measure of level of enzyme purity, is summarized in Table 1. A cursory look at Table 1 shows increasing level of PPO specific activity with the progression of each purification step.

Table 1

Specific activity of PPO extracted from three segments of *S. melongenas* and *M. sapietum* fruits at various purification steps.

Purification step	Specific activity (U/mg protein)					
	<i>S. melongenas</i>			<i>M. sapietum</i>		
	A	MS	P	A	MS	P
Centrifuged at 32000 <i>g</i> for 20 min at 4 °C	0.68±0.87	0.78±0.67	2.08±0.87	0.58±0.99	1.98±0.67	1..28±0.95
80% (NH ₄) ₂ SO ₄	2.72±1.07	2.48±0.37	5.14±0.36	1.98±0.90	3.72±0.81	2.08±1.27
Ultra-filtration	5.18±0.65	5.88±0.91	7.77±0.15	5.91±1.27	6.88±1.07	4.78±1.05
Dialyzed at 4°C	6.78±0.77	7.18±0.27	8.07±0.47	7.78±1.03	7.85±0.77	6.98±0.93
%yield	11.3±0.78	11.9±0.9.1	12.8±0.71	10.0±0.97	12.1±0.59	10.3±0.87
[protein] mg mL ⁻¹	0.012±0.7	0.017±0.9	0.025±0.7	0.010±0.9	0.018±0.5	0.011±0.8

A: Anterior; MS: Mid-section; P: Posterior. The results are means (\bar{X}) ± S.D of three ($n = 3$) determinations.

Table 2

Kinetic constants (K_m and V_{max}) of PPO extracted from three segments of *S. melongenas* and *M. sapietum* fruits

Fruit segment	<i>S. melongenas</i>		<i>M. sapietum</i>	
	K_m (mM)	V_{max} (U)	K_m (mM)	V_{max} (U)
Anterior	2.0±0.99 ^a	0.251±0.04 ^a	3.9±0.69 ^a	0.234±0.09 ^a
Mid-section	1.5±0.09 ^a	0.393±0.60 ^b	5.8±0.69 ^b	0.420±0.06 ^b
Posterior	4.6±0.49 ^b	0.602±0.09 ^c	4.9±0.11 ^{b,c}	0.241±0.05 ^{a,c}

The results are means (\bar{X}) ± S.D of three ($n = 3$) determinations. Means in the columns with the same letter are not significantly different at $p < 0.05$ according to LSD.

The kinetic parameters of PPO extracted from three segments of the two fruits are presented in Table 2. The K_m values of PPO (PPO_{K_m}) extracted from the three segments of *S. melongenas* were in the range of 1.5±0.09 - 4.6±0.49 mM. Furthermore, PPO_{K_m} of mid-section and anterior segments showed no significant difference ($p < 0.05$), whereas the posterior gave $PPO_{K_m} = 4.6±0.49$ mM; $p > 0.05$. Overall, PPO_{K_m} of the three segments of *S. melongenas* was in the order: Mid-Section < Anterior < Posterior. PPO maximum activity ($PPO_{V_{max}}$) was highest in the posterior segment ($PPO_{V_{max}} = 0.602±0.09$ U) compared with the other two segments: mid-section $PPO_{V_{max}} = 0.393±0.60$ U; $p > 0.05$ and anterior $PPO_{V_{max}} = 0.251±0.04$ U; $p > 0.05$.

PPO extracted from the mid-section of *M. sapietum* exhibited the highest K_m value ($PPO_{K_m} =$

5.8±0.69 mM) compared with the anterior ($PPO_{K_m} = 3.9±0.69$ mM) ($p > 0.05$) and posterior ($PPO_{K_m} = 4.9±0.11$ mM) segments ($p < 0.05$). *M. sapietum* anterior $PPO_{V_{max}}$ was not significantly different ($p < 0.05$) from posterior $PPO_{V_{max}}$. *M. sapietum* mid-section $PPO_{V_{max}}$ was highest compare to other two segments ($p > 0.05$). An overview of Table 2 shows that *M. sapietum* PPO_{K_m} values were relatively higher than those of *S. melongenas*. Posterior *S. melongenas* exhibited the highest $PPO_{V_{max}} = 0.602±0.09$ U, whereas the lowest value was registered in the anterior segment of *M. sapietum* $PPO_{V_{max}} = 0.234±0.09$ U.

Substrate specificity for PPO (PPO_{V_{max}/K_m}) extracted from various segments of the two fruits was in the range of 0.049 - 0.262 U/mM. For *S. melongenas* enzyme extract, the increasing order of PPO_{V_{max}/K_m} was Mid-Section > Posterior >

Anterior, whereas that of *M. sapietum* was Mid-Section > Anterior > Posterior. PPO_{V_{max}/K_m} between the two fruits showed strong positive correlation ($r = 0.862339$).

Discussion

The enzyme extracts from the three segments of *S. melongenas* and *M. sapietum* exhibited PPO activity, which was in conformity with previous reports elsewhere (Chikezie et al., 2007; Ünal, 2007; Queiroz et al., 2008). The present kinetic study showed that in all cases, $PPO_{S. melongenas}$ and $PPO_{M. sapietum}$ conformed to Michaelis-Menten kinetics, exhibiting different values of kinetics parameters. In agreement with the present findings, Rocha et al. (2001), had earlier noted that PPO isolated from higher plants oxidized a wide range of monophenols and *o*-diphenols with highly variable V_{max} and K_m values.

A measure of affinity of the enzyme for its substrate is defined by the K_m value. Overall, PPO extracts from *S. melongenas* exhibited higher affinity for the experimental substrate (catechol) than those extracted from *M. sapietum*. The affinity of plant PPO for the phenolic substrates was generally low (high K_m values, 2 ± 6 mM) according to Nicolas et al., (1994). Likewise, the results presented here showed that PPO_{K_m} extracted from the three segments of *S. melongenas* and *M. sapietum* was in the range of 1.5 ± 0.09 - 5.8 ± 0.69 mM (Table 2). The variability of PPO_{K_m} in the three segments of the two fruits confirmed differences in affinity of the enzymes for phenolic substrates. According to Altunkaya and Gökmen, (2011), the variability in PPO_{K_m} is diagnostic of isoenzymic forms of PPO in *Lactuca sativa*. They noted that substrate specificity in terms of V_{max}/K_m values of two fractions of PPO extracts (PPO_1 and PPO_4) was different and therefore, order of affinity of the isoenzymes for various substrates varied. Furthermore, Marshall et al. (2000) averred that variations in K_m values of *Mangifera indica* fruit extracts with concomitant difference in affinity between mono- and polyphenol substrates for the enzyme was an indication of the presence of isoenzyme in *M. indica* fruits. In another study, Cornish-Bowden and Cardenas (2010), in their research paper showed that variability in kinetic

Table 3
Catechol specificity for PPO extracted from three segments of *S. melongenas* and *M. sapietum* fruits

Fruit Segment	V_{max}/K_m (U/mM)	
	<i>S. melongenas</i>	<i>M. sapietum</i>
Anterior	0.126	0.060
Mid-section	0.262	0.072
Posterior	0.131	0.049

parameters of non-Michaelis-Menten enzymes provided necessary information for analyzing metabolic pathways associated with isoenzymes. Values of PPO_{K_m} of the mid-section and posterior segments of *S. melongenas* showed significant difference ($p > 0.05$) whereas, the difference in K_m values of *M. sapietum* enzyme extract between the mid-section and posterior segments was not significant ($p < 0.05$) (Table 2). These observations indicated the presence of isoenzymic forms of PPO in the corresponding segments of the two fruits as reported elsewhere (Vámos-Vigyázó, 1981; Barthet, 1997; Casado-Vela et al., 2005; Chikezie, 2006; Altunkaya and Gökmen, 2011). In another perspective, the K_m values could also give an insight into the physiologic concentrations of the PPO substrates in the three portions of the two fruits under investigation. More than four decades ago, Sheen, (1969) posited that there is correlation between phenolic quantity and oxidase activity, which varied depending upon the organs and tissues. However, it is worthy to note here that the experimentally observed K_m value is a function of pH and ionic strength of the enzyme assay solution (Rivas and Whitaker, 1973; Janovitz-Klapp et al., 1989; Valero and García-Carmona, 1998; Nicolas et al., 1994). The variability of $PPO_{V_{max}}$ in the various segments of the two fruit enzyme extracts was a pointer to the fact that differences exist in the relative abundance and distribution of PPO in biologic tissues and systems (Qudsieh et al., 2002; Sirhindi, 2003; Escobar et al., 2008; Zamorano et al., 2009).

PPO_{V_{max}/K_m} defines the suitability of the experimental substrate (catechol) for PPO extracted from the two fruits. An overview of

Table 3 shows that catechol exhibited relatively low specificity for PPO_{M. sapientum} compared to PPO_{S. melongenas}. Previous reports have established that certain categories of phenolic compound are poor substrate to PPO by virtue of their specificity ratio V_{max}/K_m (Rocha et al., 2001; Fortea et al., 2009). For instance, monophenol (tyrosine) was found to be a poor substrate for the apple PPO (Nicolas et al., 1994; Rocha et al., 2001). Richard-Forget et al. (1992) showed that several compounds such as chlorogenic acid and catechins appeared to be better substrates than 4-methylcatechol for PPO extracted from Red Delicious apples. The kinetic parameters (Tables 2 and 3) indicated that catechol was a better substrate for PPO_{S. melongenas} than PPO_{M. sapientum}. Furthermore, the experimentally observed kinetic parameters of *S. melongenas* and *M. sapientum* signified the presence of PPO isoenzyme and non-uniform distribution of PPO in the two fruits.

References

- Altunkaya, A.** and **V. Gökmen.** 2011. 'Purification and characterization of polyphenol oxidase, peroxidase and lipoxygenase from freshly cut Lettuce (*L. sativa*)'. *Food Technology and Biotechnology.* 49 (2): 249–256.
- Anderson, J. V., E. P. Fuerstb, W. J., Hurkman, W. H. Venselc, and C. F. Morrisd.** 2006. Biochemical and genetic characterization of wheat (*Triticum spp.*) kernel polyphenol oxidases. *Journal of Cereal Science.* 44(3): 353–367.
- Anthon, G. E.** and **D. M. Barrett.** 2002. Kinetic parameters for the thermal inactivation of quality-related enzymes in Carrots and Potatoes. *Journal of Agriculture and Food Chemistry.* 50(14): 4119-4125.
- Barthet, V. J.** 1997. Polyphenol oxidases from cassava (*Manihot esculenta* C.) root: extraction, purification and characterization.(Abstract).http://digitool.Library.McGill.CA:80/R/-?func=dbin-jumpfull&object_id=34697&silolibrary=GEN01. Accessed 30th January, 2013.
- Beena, P.** and **L. R. Gowda.** 2000. Purification and characterization of polyphenol oxidase from the seeds of field bean. *Journal of Agricultural and Food Chemistry,* 48(9): 3839-46.
- Bradford, M. M.** 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein–dye binding. *Annal of Biochemistry.* 72:248–254.
- Broothaerts, W., J. McPherson, B. Li, E. Randall, W.D. Lane and P.A. Wierma.** 2000. Fast apple (*Malus x domestica*) and tobacco (*Nicotiana tobacum*) leaf polyphenol oxidase activity assay for screening transgenic plants. *Journal of Agriculture and Food Chemistry.* 48: 5924-5928.
- Casado-Vela, J., S. Selles and R. Bru.** 2005. Purification and kinetic characterization of polyphenol oxidase from tomato fruits (*Lycopersicon esculentum* cv. *Muchamiel*). *Journal of Food Biochemistry.* 29: 381–401.
- Chikezie, P. C.** 2006. Extraction and activity of polyphenoloxidase from kolanuts (*Cola nitida* and *Cola acuminata*) and cocoa (*Theobroma cacao*). *Journal of Agriculture and Food Sciences.* 4(2):115-124.
- Chikezie, P. C., B. A. Amadi and C. M. Chikezie.** 2007. Evaluation of two polyphenoloxidase kinetic parameters at three stages of *Musa sapientum* fruit maturation. *International Journal of Agriculture and Food Systems.* 1(1): 63-67.
- Chisari, M., R. N. Barbagallo and G. Spagna.** 2007. Characterization of polyphenol oxidase and peroxidase and influence on browning of cold stored Strawberry fruit. *Journal of Agriculture and Food Chemistry.* 55(9): 3469-3476.
- Cornish-Bowden, A.** and **M. L. Ca'rdenas.** 2010. Specificity of non-Michaelis-Menten enzymes: necessary information for analyzing metabolic pathways. *Journal of Physical Chemistry B,* 114(49): 16209–16213.
- Escobar, M. A., A. Shilling, P. Higgins, S. L. Uratsu and A. M. Dandekar.** 2008. Characterization of polyphenol oxidase from Walnut. *Journal of the American Society for Horticultural Science.* 133(6): 852-858.
- Fawzy, A. M.** 2005. Purification and some properties of polyphenol oxidase from Apple (*Malus Domestica* Borkh.). *Minia Journal of*

Agricultural Research and Development. 25: 629-644.

- Fortea, M. I., S. López-Miranda, A. Serrano-Martínez, J. Carreño and E. Núñez-Delgado.** 2009. Kinetic characterisation and thermal inactivation study of polyphenol oxidase and peroxidase from table grape (Crimson Seedless). *Food Chemistry*. 113 (4): 1008–1014.
- Fujita, S., N. B. Saari, M. Maegawa, T. Tetsuka, N. Hayashi and T. Tono.** 1995. Purification and properties of polyphenol oxidase from cabbage. *Journal of Agricultural and Food Chemistry*, 43: 1138-1142.
- Galeazzi, M. A. M., V. C. Sgarbieri and S. M. Constantinides** 1981. Purification and physicochemical characterization of polyphenol oxidase from a dwarf variety of banana. *Journal of Food Science*, 46(1): 150-155.
- Gandía-Herrero, F., F. García-Carmona and J. Escribano.** 2004. Purification and characterization of a latent polyphenol oxidase from beet root (*Beta vulgaris* L.). *Journal Agriculture and Food Chemistry*. 52(3): 609-615.
- Gouzi, H., T. Coradin, E. M. Delicado, M. U. Ünal, and A. Benmansour.** 2010. Inhibition kinetics of *Agaricus bisporus* (J.E. Lange) Imbach polyphenol oxidase *The Open Enzyme Inhibition Journal*, 3: 1-7.
- Janovitz-Klapp, A. H., F. C. Richard-Forget and J. J. Nicolas.** 1989. Polyphenoloxidase from apple, partial purification and some properties. *Phytochemistry*. 28: 2903-2907.
- Jiménez-Atiéndar M., P. M. Angeles and F. García-Carmona.** 1991. Activation of polyphenol oxidase by polyamines. *Biochemistry International*. 25(5): 861-868.
- Kavrayan, D. and T. Aydemir.** 2001. Partial purification and characterization of polyphenol oxidase from peppermint (*Mentha piperita*). *Journal of Food Chemistry*, 74(2):147-154.
- Klabunde, T., C. Eicken, J. C. Sacchettini and B. Krebs.** 1998. Crystal structure of a plant catechol oxidase containing a dicopper center. *Nature Structural Biology*. 5(12): 1084–1090.
- Kumar, V. B. A., T. C. K. Mohan and K. Murugan.** 2008. Purification and kinetic characterization of polyphenol oxidase from Barbados cherry (*Malpighia glabra* L.). *Food Chemistry*. 110 (2): 328–333.
- Laveda, F., E. Nunez-Delgado, F. Garcia-Carmona and A. Sanchez-Ferrer.** 2001. Proteolytic activation of latent Paraguaya peach PPO. Characterization of monophenolase activity. *Journal Agriculture and Food Chemistry*. 49(2): 1003-1008.
- Lineweaver, H. and D. Burk.** 1934. The determination of enzyme dissociation constants. *Journal of the American Chemical Society*. 56: 658–666.
- Madani, I., P. M. Lee and L. K. Hung.** 2011. Partial purification and characterisation of polyphenol oxidase from *Hibiscus rosa-sinensis* L. *2nd International Conference on Biotechnology and Food Science*. Singapore.
- Marques, L., A. Fleuriet, J. C. Cleyet-Marel and J. Macheix.** 1994. Purification of an apple polyphenol oxidase isoform resistant to SDS-proteinase K digestion. *Phytochemistry*, 36: 1117–1121.
- Marshall, M. R., J. Kim and C. Wei.** 2000. Enzymatic browning in fruits, vegetables and sea foods. *Food and Agricultural Organization*, 41:259-312.
- Mayer, A. M.** 2006. Polyphenol oxidases in plants and fungi: Going places? A review. *Phytochemistry*, 67(21): 2318–2331.
- Nagai, T. and N. Suzuki.** 2001. Partial purification of polyphenol oxidase from Chinese cabbage (*Brassica rapa* L). *Journal of Agriculture and Food Chemistry*, 49(8): 3922-3926.
- Nicolas, J. J., F. C. Richard-Forget, P. M. Goupy, M. J. Amiot and S. Y. Aubert.** 1994. Enzymatic browning reactions in apple and apple products. *Critical Reviews in Food Science Nutrition*, 34(2): 109-157.
- Okot-Kotber, M., A. Liavoga, K. J. Yong and K. Bagorogoza.** 2002. Activation of polyphenol oxidase in extracts of bran from several wheat (*Triticum aestivum*) cultivars using organic solvents, detergents, and chaotropes. *Journal Agriculture and Food Chemistry*. 50(8): 2410-2417.
- Oktay, M., I. Kufrevioglu, I. Kocacaliskan and H. Sakiroglu.** 1995. Polyphenol oxidase from Amasya apple. *Journal of Food Science*, 60: 494-496.

- Parkin, K. L.** 2008. *Enzymes*. In: Damodaran, S., Parkin, K. L and Fennema, O. R (eds.). Fennema's Food Chemistry. 4th ed. CRC Press, New York.
- Prohp, T. P., K. E. Ekpo, E. V. Osagie, A. Osagie and H. Obi.** 2009. Polyphenol contents and polyphenol oxidase activities of some Nigerian kolanuts. *Pakistan Journal of Nutrition*. 8(7): 1030-1031.
- Qudsieh, H. Y., S. Yusof, A. Osman and R. A. Rahman.** 2002. Effect of maturity on chlorophyll, tannin, color and polyphenol oxidase (PPO) activity of sugarcane juice (*Ssaccharum officinarum* var. Yellow cane). *Journal Agriculture and Food Chemistry*. 50: 1615–1618.
- Queiroz, C., L. Mendes, L. Maria, E. Fialho, M. Valente and L. Vera.** 2008. Polyphenol oxidase: characteristics and mechanisms of browning control. *Food Reviews International*. 24 (4): 361-375.
- Richard-Forget, F. C., M. A. Rouet-Mayer, P. M. Goupy, J. Philippon and J. J. Nicolas.** 1992. Oxidation of chlorogenic acid, catechins, and 4-methylcatechol in model solutions by apple polyphenol oxidase. *Journal of Agricultural and Food Chemistry*, 40: 2114-2122.
- Rivas, N. and J. R. Whitaker.** 1973. Purification and some properties of two polyphenol oxidases from Bartlett Pears. *Plant Physiology*. 52(5): 501-507.
- Rocha, A. M. C. N. and A. M. M. B. Morais.** 2001. Characterization of polyphenoloxidase (PPO) extracted from Jonagored apple. *Food Control*, 12(2): 85-90.
- Santosh, R. K., P. Beena, A. G. Appu and L. R. Gowda.** 2006. The conformational state of polyphenol oxidase from field bean (*Dolichos lablab*) upon SDS and acidpH activation, *Biochemistry Journal*, 395(3):551-562.
- Sélles-Marchart, S., J. Casado-Vela and R. Bru-Martínez.** 2006. Isolation of a latent polyphenol oxidase from loquat fruit (*Eriobotrya japonica* Lindl.): Kinetic characterization and comparison with the active form. *Archive of Biochemistry and Biophysics*. 446(2): 175–185.
- Sheen, S. J.** 1969. The distribution of polyphenols, chlorogenic acid oxidase and peroxidase in different plant parts of tobacco, *Nicotiana tabacum* L *Phytochemistry*. 8 (10): 1839–1847.
- Sirhindi, G.** 2003. Polyphenol oxidase production and activity under high temperature exotherm in vegetative organs of *Thuja orientalis*. *Environmental Informatics Archives*. 1: 574-580.
- Thipyapong, P., M. D. Hunt and J. C. Steffens.** 2004. Antisense down regulation of polyphenol oxidase results in enhanced disease susceptibility. *Planta*. 220: 105–107.
- Ünal, M. U.** 2007. Properties of polyphenol oxidase from Anamur banana (*Musa cavendishii*). *Food Chemistr*, 100 (3):909–913.
- Valero, E. and F. Garcia-Carmona.** 1998. pH-dependent effect of sodium chloride on latent grape polyphenol oxidase. *Journal of Agriculture and Food Chemistry*, 46 (7): 2447-2451.
- Vamos-Vigyazo, L.** 1981. Polyphenoloxidase and peroxidase in fruits and vegetables. *Critical Review in Food Science and Nutrition*, 15(1): 49-127.
- Yağar H. and A. Sağıroğlu .**2002. Partially purification and characterization of polyphenol oxidase of quince. *Turkish Journal of Chemistry*, 26: 97- 104.
- Zamoranoa, J., O. Martínez-Álvarezb, P. Monterob and M. Gómez-Guillénb.** 2009. Characterisation and tissue distribution of polyphenol oxidase of deep water pink shrimp (*Parapenaeus longirostris*). *Food Chemistry*. 112 (1): 104–111.